Arylcarbamate Derivatives of 1-Piperidineethanol as Potent Ligands for 5-HT₄ Receptors

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A series of carbamate derivatives (7) of 2-(1-piperidinyl)ethyl 4-amino-5-chloro-2-methoxybenzoates, which have been described as potent agonists and antagonists of 5-HT₄ receptors, were synthesized. They were evaluated using radioligand binding assays with [³H]GR 113808, a 5-HT₄ receptor selective ligand, in the rat striatum and the electrically stimulated myenteric plexus longitudinal muscle of the guinea pig. In contrast to the previously described ester derivatives, a drop in the affinity for 5-HT₄ receptors was observed and the compounds were inactive as agonists in the guinea pig ileum preparation. Unexpectedly, the *ortho*-substituted carbamates **8b**, **c** (R' = H, RO = MeO or EtO, R'' = H) had nanomolar affinity for 5-HT₄ receptors ($K_i = 8.9 \pm 0.5$ and 2.6 ± 0.4 nM, respectively). As reported previously, the *cis*- or *trans*-3,5dimethyl substitution of piperidine (**8n**,**o**) was particularly favorable ($K_i = 1.1 \pm 0.6$ nM for both isomers). **8c** is an antagonist equipotent to the 5-HT₄ receptor antagonist SDZ 205-557 (1).

Stimulation of 5-HT₄ receptors is responsible for the gastrokinetic effects of drugs such as the benzamides.¹⁻³ In the guinea pig^{4b} they are located on the ileal enteric nerve where they mediate the release of acetylcholine, thus bringing about the first phase of the muscle contraction-concentration curve to 5-HT. The effect of stimulation of 5-HT₄ receptors seems to be speciesdependent as very different results have been obtained in the rat where 5-HT₄ receptor stimulation in the rat ileum induced relaxation via an atropine-insensitive mechanism, suggesting a localization¹ on the smooth muscle. On the other hand, in man 5-HT₄ receptormediated effects have not been observed in the ileum. However 5-HT₄ receptors are present in the colon¹ where they appear to mediate a direct effect on the smooth muscle, inducing muscle relaxation. Several studies have shown the presence of 5-HT₄ receptors in the stomach of a variety of species, such as the guinea pig, rat, dog, and man,¹ where they mediate an increase in gastric emptying. 5-HT₄ receptors are also responsible for the relaxation of the esophagus muscle in the rat.4a

Considerable progress has been made in the localization and study of the functions of 5-HT₄ receptors with the development of potent and selective antagonists such as SDZ 205-557 (1),⁵ SB 204070A (2),^{6.7} SB207266 (3),⁸ and GR 113808 (4)⁹ and, in particular, by the use of a number of them as radiolabeled ligands.^{7,9,10}

Research into selective 5-HT₄ receptor agonists as gastrointestinal (GI) prokinetic drugs for the treatment of motility disorders such as gastroesophageal reflux, functional dyspepsia, and constipation has received considerable interest recently. Many of the compounds described as possessing these properties, such as cisapride, renzapride, SC-53116 (5), BRL 24682 (6), and zacopride, are members of the generic benzamide family being amides of 4-amino-5-chloro-2-methoxybenzoic acid,¹ which appears to be an important structural requirement for 5-HT₄ receptor agonist activity.

Recently, a new family of esters, derived from 2-(1piperidinyl) 4-amino-5-chloro-2-methoxybenzoate,¹¹ were described by us, as potent selective ligands for 5-HT₄ receptors. Almost all of these derivatives were potent agonists for this receptor, although it was shown that the introduction of a 3,5-dimethylpiperidine moiety in the basic chain brought about a dramatic change in the pharmacological profile, as 2-(3,5-dimethylpiperidino)ethyl 4-amino-5-chloro-2-methoxybenzoate¹² was demonstrated to be a potent 5-HT₄ receptor antagonist. However, the presence of the ester function is an important drawback for their administration in vivo due to their putative short half-lives. To overcome this problem, a series of carbamates (7) derived from the esters previously described were synthesized, several structural variations of the aromatic moiety were carried out (compounds 8), and the role of the oxygen atom in the basic chain was evaluated with the synthesis of the amidic derivative 9.

Chemistry

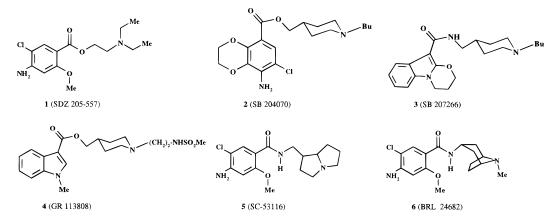
The carbamates 7, structurally closely related to benzoates derived from 4-amino-5-chloro-2-methoxybenzoic acid, were prepared according to the synthetic pathway described in Scheme 1. 5-Chloro-o-anisidine was acetylated and transformed into the corresponding nitro amine derivative 10 according to the process described by Hadley.¹³ It was treated with phosgene in a toluene solution to give 5-chloro-2-methoxy-4nitrophenyl isocyanate (11) which reacted with the different amino alcohols, easily synthesized by the reaction of 2-bromoethanol and the secondary amines in the presence of NaOH, to give compounds 12. The nitro group of compounds 12 was reduced into the amino derivatives 7a-g by the reaction of Na₂S₂O₄ in a MeOH/ H_2O^{14} mixture with a moderate yield. Compounds were isolated as hydrochloride or oxalate salts.

Mono- or disubstituted carbamates **8** were prepared from the isocyanates **13** which were either commercially

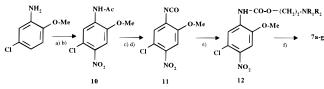
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Chart 1



Scheme 1^a

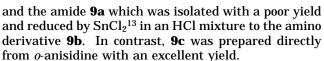


 a (a) AcCl, CH₂Cl₂, rt; (b) HNO₃, H₂SO₄, AcOH; (c) NaOH, EtOH/ H₂O, reflux; (d) 20% toluene solution of COCl₂, CH₂Cl₂, 0 °C; (e) R₂R₁N(CH₂)₂OH, 0 °C, reflux; (f) Na₂S₂O₄, MeOH/H₂O.

available or prepared from the corresponding anilino derivatives by reaction with phosgene. Their condensation (Scheme 2) with the corresponding amino alcohols in THF under reflux gave compounds **8a–i,m–o. 8j–l** were obtained by the reduction of the corresponding nitro derivatives **8p–r** by Na₂S₂O₄.¹⁴ The derivatives of 3,5-dimethylpiperidine (compounds **7g**, **8m–o**) were obtained as a 4:1 mixture of *cis* and *trans* compounds, respectively, which were separated by column chromatography and characterized by their ¹H-NMR spectra.

Amide **9b** was prepared from 5-chloro-2-methoxy-4nitroaniline (Scheme 3) which was condensed with 4-chlorobutyryl chloride to give the chloro amide **15**. The reaction of **15** (Ar = 5-chloro-2-methoxy-4-nitrophenyl) and piperidine gave a mixture of 1-(5-chloro-2-methoxy-4-nitrophenyl)-2-pyrrolidone by a cyclization reaction

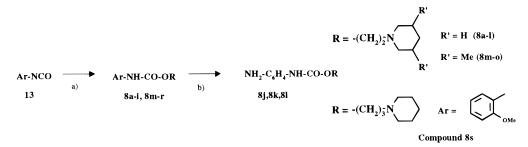
Scheme 2^a



Biological Methods

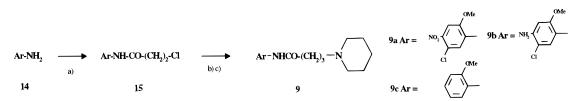
Assessment of the affinity of the compounds for 5-HT₄ receptors was performed using radioligand binding assays with [³H]GR 113808 and rat striatal membranes.¹⁵ The existence of 5-HT₄ receptors has been clearly demonstrated in the central nervous system (CNS), and these receptors have been shown to be indentical to those in the periphery, in particular, those in the intestine.¹⁶ Consequently, binding assays in the brain constitute a valuable test to select compounds which will be active in the GI tract.

The facilitatory role of 5-HT₄ receptors in the peristaltic reflex was demonstrated by Craig and Clarke⁴ who found that both 5-HT₃ and 5-HT₄ receptor agonists induced peristalsis in the guinea pig isolated ileum maintained at a subthreshold intraluminal pressure. The 5-HT₄ receptor agonist activity of the tested compounds was measured in the electrically stimulated myenteric plexus and longitudinal muscle of the guinea pig ileum, and it was assessed as the concentration (EC₅₀, nM) which gave a 50% increase in the response



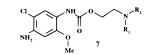
^{*a*} (a) ROH, 0 °C, reflux; (b) Na₂S₂O₄, MeOH/H₂O, only with the compounds **8p**-**r**.

Scheme 3^a



^{*a*} (a) ClCO-(CH₂)₃-Cl, benzene, rt; (b) piperidine, toluene, reflux for 4 h or piperidine, NaI, rt for 2 days; (c) 1-(3-(1-piperidinyl)propanecarboxamido)-4-nitro-5-chloro-2-methoxybenzene, SnCl₂, HCl.

Table 1. Pharmacological Activity of the Carbamates 7 at5-HT4 Receptors



		Binding assays	Functional activity at 5-HT ₄ receptors		
Compd	NR_1R_2	Ki, nM ^a , 5-HT₄	EC50, nMb	IC ₅₀ , nM ^C	
7a	-x	39 ± 7.2	>1000	ned	
7ь	-x	125 ± 67	~200 (23%)	ne	
7c	-N	14.5 ± 6.4	100 (30%)	207[176-244]	
7d		150 ± 68	300 (39%)	1000	
7e	-x	10.2 ± 5.5	NTe	NT	
7f		25.1 ± 4.8	~60 (25%)	200	
7g		2.4 ± 0.5	>1000	180	
$\underset{O-Me}{\overset{Cl}{\underset{0}{}}} \underset{O-Me}{\overset{CO-O-(CH_{2})_{T}}} N \underset{Me}{\overset{Me}{\underset{Me}{}}}$		0.26 ± 0.05	Įf	11 [9-14]	
GR 113808 (4) ML 10302 SDZ 205-557 ($\begin{array}{c} 0.15 \pm 0.03 \\ 1.07 \pm 0.5 \\ 5.5 \pm 0.7 \end{array}$	I 4 [3.7-4.3] (78%) I	13 [9-18] ne 77 [55-130]	

 a [³H]GR 113808 was used as the radioligand in the rat striatum; $K_i \pm SEM$ values were determined from the Cheng–Prussof equation. b The agonist activity was assessed as the concentration which gave a 50% increase in the response to electrical stimulation (EC_{50}) in the guinea pig ileum and is expressed as a percent of the maximum 5-HT response; 95% confidence limits are in brackets. c The antagonist activity (IC_{50}) was calculated as the concentration which produced a 50% reduction in the response to 5-HT in the guinea pig ileum; 95% confidence limits are in brackets. d ne = not capable of evaluation. e NT = not tested. f I = inactive up to 10^{-5} M.

to electrical stimulation with regard to its maximum and related to the maximal effect of 5-HT (100%). Antagonist activity (IC₅₀, nM) was calculated as the concentration which produced a 50% reduction in the submaximal (80%) of 5-HT-induced contractions. The activity of the compounds was compared to those of the reference compounds: ML 10302 (2-(1-piperidinyl)ethyl 4-amino-5-chloro-2-methoxybenzoate),¹¹ 2-(*cis*-3,5-dimethylpiperidino)ethyl 4-amino-5-chloro-2-methoxybenzoate,¹² a recently described potent antagonist, and compounds 1 and 4.

Results and Discussion

The results in Table 1 show that the carbamates **7** are not as good ligands for 5-HT₄ receptors as the corresponding esters described previously;^{11b} in particular, **7a** ($K_i = 39$ nM) was clearly less potent than ML 10302. However, as had already been noted,¹² an increase in the affinity for the receptor was observed following methyl substitution, particularly with the methylpiperidine derivatives **7e**–**g**. The favorable role of the 3,5-dimethyl substitution was again noted for **7g** which had nanomolar affinity. No compound displayed significant agonist activity by increasing the response to electrical stimulation in the guinea pig ileum. Only compounds **7c**,**f**,**g** produced a moderate inhibition of the 5-HT response in the guinea pig ileum. These data

show that, in contrast to the esters, the carbamate function is less favorable for recognition by the 5-HT₄ receptor and the basic moiety does not influence the pharmacological profile.

In the benzamide series derived from 4-amino-5chloro-2-methoxybenzoic acid, no clear structure-activity relationships have been described for the role of aromatic substituents on pharmacological activity. Preliminary studies on ML 10302 showed that such substituents were essential for maintaining affinity for 5-HT₄ receptors, as a large drop in the affinity value was observed with any mono- or disubstituted benzoate esters of 2-piperidinoethanol.¹⁷ Different results were obtained with the monosubstituted carbamates 8. The data reported in Table 2 show the favorable influence of monosubstitution by MeO or EtO in the ortho position (compounds **8b**, **c**, **f**, **m**, **n**), while substitution in the *meta* or para position brought about a large decrease in the biological activity (compounds 8d,e). The presence of the amino function was particularly harmful and could explain the decrease in activity observed with the carbamates 7 with regard to the corresponding esters. Only methoxy or ethoxy substitution in the ortho position had a favorable effect, as NH₂, Cl, CN,¹⁸ and NO_2^{18} substitutions produced inactive compounds ($K_i >$ 1000 nM) in the binding assays. On the other hand, the length of the basic chain is an important structural parameter in recognition by the binding site because the carbamate **8s** ($K_i = 81$ nM), derived from 3-(1-piperidinyl)propanol, was 1 order of magnitude less potent than the corresponding compound **8b** ($K_i = 8.9$ nM). As observed previously, the introduction of 3,5-dimethylpiperidine was favorable, and compounds 8m-o had nanomolar affinity, the effect of the cis or trans stereochemistry not being very marked.

In contrast to the esters, no compound displayed agonist activity in the functional assay for 5-HT₄ receptors. Thus, compounds **8b**,**c**,**f** were antagonists in the guinea pig ileum, and **8c** was equipotent to **1**. An increase in the inhibitory effect was not observed with the 3,5-dimethyl derivatives (compounds **8m**–**o**) which were equipotent to or less potent than compound **8c**.

In a previous paper,¹² we emphasized the important role of the oxygen atom of the basic chain in the recognition of the 5-HT₄ receptor binding site by the benzoate series. This finding was confirmed by the results obtained with the amidic derivatives **9b**,**c** which were weak ligands for 5-HT₄ receptors ($K_i > 1000$ nM).

In summary, the present study has shown that the carbamate derivatives of 1-piperidineethanol and 2-(3,5dimethylpiperidino)ethanol are potent ligands for 5-HT₄ receptors. It emphasized the importance of the aromatic substituents and, particularly, of *o*-methoxy or *o*-ethoxy substituents for recognition by the 5-HT₄ receptor. The results demonstrated, unexpectedly, the unfavorable role of the amino group in the 4-amino-5-chloro-2methoxybenzoic acid pharmacophore. Unlike the corresponding esters, no potent agonist was found with an unsubstituted piperidine ring, suggesting the inability of such molecules to occupy the putative agonist site suggested previously.¹² Several investigations are in course to understand the cause of the weak functional activity of these compounds on the GI receptors and their potent affinity for central 5-HT₄ receptors.

compd		R′		functional activity at 5-HT4 receptors	
	Ar		binding assays <i>K</i> _i , nM, ^a 5-HT ₄	EC_{50} , nM^b	IC_{50} , nM^c
8a	C ₆ H ₅	Н	31.7 ± 7.5	>1000	~1000
8b	2-OMeC ₆ H ₄	Н	8.9 ± 0.5	${\sim}100$ (14%)	${\sim}300$
8c	$2-OEtC_6H_4$	Н	2.6 ± 0.4	>1000	62 [40-95]
8d	3-OMeC ₆ H ₄	Н	469 ± 44	\mathbf{NT}^d	NT
8e	4-OMeC ₆ H ₄	Н	>1000	NT	NT
8f	2-OMe-5-ClC ₆ H ₃	Н	4.2 ± 0.5	${\sim}1000$ (14%)	30 (60%)
8g	$2-ClC_6H_4$	Н	>1000	NT	NT
8 h	3-ClC ₆ H ₄	Н	60.8 ± 5.1	>1000	>1000
8i	$4-ClC_6H_4$	Н	728 ± 63	NT	NT
8j	$2-NH_2C_6H_4$	Н	>1000	NT	NT
8ĸ	3-NH ₂ C ₆ H ₄	Н	>1000	NT	NT
81	$4-NH_2C_6H_4$	Н	>1000	NT	NT
8m	2-OMeC ₆ H ₄	Me, cis	2.6 ± 0.52	>1000	160 (60%)
8n	$2-OEtC_6H_4$	Me, cis	1.05 ± 0.68	>1000	30 (70%)
80	$2-OEtC_6H_4$	Me, trans	1.16 ± 0.6	>1000	100

a - d See corresponding footnotes in Table 1.

Experimental Section

Chemistry. Melting points were determined on a Mettler FP 61 melting point apparatus and are uncorrected. ¹H- and ¹³C-NMR spectra were recorded on a Bruker AC 200 spectrometer at 200 and 50 MHz, respectively. Chemical shifts are reported in parts per million (δ) relative to tetramethylsilane as the internal standard, and signals are quoted as s (singlet), ds (dedoublet singlet), d (doublet), dd (dedoublet doublet), t (triplet), dt (dedoublet triplet), q (quartet), br s (broad singlet), or m (multiplet). Elemental analyses were performed at the CNRS's analysis services in Châtenay-Malabry (France) and were within 0.4% of the theoretical values unless otherwise noted.

Materials. Tetrahydrofuran (THF) was distilled from sodium/benzophenone. Acetonitrile, dimethylformamide, toluene, and the usual solvents were purchased from SDS (Paris, France). Column chromatography was performed on Merck silica gel 60 (70/230 mesh). Thin-layer chromatography was done on silica gel 60F-254 (0.26 mm thickness) plates.

4-Amino-5-chloro-2-methoxybenzoic acid, 2-bromoethanol, the mixture of *cis*- and *trans*-3,5-dimethylpiperidine, hexamethylpiperidine, 4-methylpiperidine, 3-methylpiperidine, 2-methylpiperidine, pyrrolidine, 1-piperidineethanol, 1-pyrrolidineethanol, and *N*,*N*-diethylaminoethanol were purchased from Aldrich (France). 2-(Hexamethyleneimino)ethanol, 2-(*cis*, *trans*-3,5-dimethylpiperidino)ethanol, 2-(4-methylpiperidino)ethanol, 2-(3-methylpiperidino)ethanol, and 2-(2-methylpiperidino)ethanol were prepared from 2-bromoethanol according to the process already reported.¹⁹ *N*-Acetyl-5-chloro-4-nitro-2-anisidine was synthesized from 5-chloro-2-anisidine according to the process reported by Hadley.¹³

5-Chloro-4-nitro-2-anisidine. *N*-Acetyl-5-chloro-4-nitro-2-anisidine was refluxed with stirring for 2 h in a solution of 10% NaOH in an EtOH/H₂O mixture (7:3) to give, after evaporation and extraction with CH₂Cl₂, 5-chloro-4-nitro-2-anisidine which was recrystallized in an acetone/AcOEt mixture (68%) as yellow spangles: ¹H NMR (CDCl₃) δ 7.57 (s, 1H), 6.69 (s, 1H), 4.53 (m, 2H), 3.91 (s, 3H).

General Method for the Preparation of Carbamates 7a–g and 8j–1. 5-Chloro-4-amino-2-anisidine or another appropriate aniline (5 mmol) was added to a solution of phosgene (2.74 mL, 20% in toluene) in anhydrous CH_2Cl_2 (20 mL) at 0 °C under argon. The solution was stirred for 15 min, and NEt₃ (1.6 mL, 11.5 mmol) was added. After 15 min, a solution of the amino alcohol (1.3 mL, 10 mmol) in anhydrous CH_2Cl_2 (5 mL) was added dropwise at 0 °C. The reaction mixture was stirred for 10 min and refluxed overnight. The solvent was evaporated under reduced pressure, and the

residue was taken up with 10% aqueous NaHCO3 and extracted with CH_2Cl_2 (3 \times 30 mL). The combined organic extracts were dried over MgSO₄ and concentrated in vacuo. The residue was chromatographed on a column of silica gel with a CH₂Cl₂/MeOH mixture (9:1) to give a crude product which was reduced to the amino derivative according to the following procedure. It was dissolved in a mixture of MeOH (30 mL), H_2O (25 mL), and 1.92 g of $Na_2S_2O_4$, and the solution was stirred for 4 h at room temperature (7d-f, 8j-l) or at 60 °C (7a,b,g). The solvent was removed by distillation in vacuo, and the residue was taken up with brine and extracted with CH_2Cl_2 (4 \times 25 mL). The aqueous layer was made alkaline with a 1 N NaOH aqueous solution and extracted again with CH_2Cl_2 (4 \times 25 mL). The combined organic layers were dried over Na₂SO₄ and concentrated to give the amino derivative which was transformed into the hydrochloride salt with a 2 N HCl methanol solution or the oxalate salt by treatment with oxalic acid in acetone. The salt was recrystallized according to the usual procedure.

General Method for the Preparation of Carbamates 8b–e,g–i,n–s. These compounds were prepared from the corresponding anilines according to the previous procedure except for the nitro reduction step and were transformed into the hydrochloride or oxalic salt according to classical methods.

General Method for the Preparation of Carbamates 8a,f,m. To a stirred solution of commercial isocyanate (10 mmol) in anhydrous CH_2Cl_2 was added dropwise the corresponding amino alcohol (20 mmol). The reaction mixture was refluxed for 1.5 h. After evaporation of the solvent *in vacuo*, the residue was chromatographed on a column of silica gel with a $CH_2Cl_2/MeOH$ mixture (9:1), and the compound was transformed into the hydrochloride salt with a 2 N HCl methanol solution or the oxalate salt by treatment with oxalic acid in acetone. The salt was recrystallized according to the usual procedure.

2-Piperidinoethyl *N*-(4-amino-5-chloro-2-methoxyphenyl)carbamate (7a): hydrochloride salt (11%) from *i*-Pr₂O/ MeOH; mp 196–198 °C dec; ¹H NMR (CD₃OD) δ 8.12 (s, 1H), 7.03 (s, 1H), 4.53 (m, 2H), 3.92 (s, 3H), 3.64 (m, 2H), 3.47 (m, 2H), 3.03 (m, 2H), 2.1–1.4 (m, 6H). Anal. (C₁₅H₂₂ClN₃O₃· HCl·3H₂O).

2-Pyrrolidinoethyl *N*-(4-amino-5-chloro-2-methoxyphenyl)carbamate (7b): hydrochloride salt (5%) from *i*-Pr₂O/ MeOH; mp 180 °C; ¹H NMR (CD₃OD) δ 7.62 (s, 1H), 6.56 (s, 1H), 4.46 (m, 2H), 3.84 (s, 3H), 3.56 (m, 2H), 3.47 (m, 4H), 2.35–2 (m, 4H). Anal. (C₁₄H₂₀ClN₃O₃·HCl).

2-(Hexamethyleneimino)ethyl N-(4-amino-5-chloro-2methoxyphenyl)carbamate (7c): oxalate salt (10%) from *i*-Pr₂O/MeOH; mp 174–176 °C; ¹H NMR (CD₃OD) δ 7.57 (s, 1H), 6.51 (s, 1H), 4.46 (m, 2H), 3.92 (s, 3H), 3.79 (m, 2H), 3.49 (m, 2H), 3.34 (m, 4H), 1.92 (m, 4H), 1.73 (m, 4H). Anal. $(C_{16}H_{24}ClN_3O_3\cdot C_2H_2O_4).$

2-(Diethylamino)ethyl *N*-(4-amino-5-chloro-2-methoxyphenyl)carbamate (7d): oxalate salt (28%) from *i*-Pr₂O/ MeOH; mp 146–148 °C; ¹H NMR (CD₃OD) δ 7.60 (s, 1H), 6.57 (s, 1H), 4.51 (m, 2H), 3.85 (s, 3H), 3.54 (m, 2H), 3.37 (m, 4H), 1.4 (m, 6H). Anal. (C₁₄H₂₂ClN₃O₃·C₂H₂O₄).

2-(3-Methylpiperidino)ethyl *N*-(4-amino-5-chloro-2methoxyphenyl)carbamate (7e): oxalate salt (9%) from *i*-Pr₂O/MeOH; mp 166–168 °C; ¹H NMR (CD₃OD) δ 7.56 (s, 1H), 6.51 (s, 1H), 4.47 (m, 2H), 3.79 (s, 3H), 3.7–3.45 (m, 2H), 3.42 (m, 2H), 3–2.8 (m, 1H), 2.5–2.72 (m, 1H), 2.1–1.7 (m, 4H), 1.35–1.05 (m, 1H), 0.99 (d, 3H). Anal. (C₁₆H₂₄ClN₃O₃· C₂H₂O₄).

2-(4-Methylpiperidino)ethyl *N*-(4-amino-5-chloro-2methoxyphenyl)carbamate (7f): oxalate salt (30%) from *i*-Pr₂O/MeOH; mp 160 °C; ¹H NMR (CD₃OD) δ 7.58 (s, 1H), 6.52 (s, 1H), 4.46 (m, 2H), 3.8 (s, 3H), 3.75–3.52 (m, 2H), 3.52– 3.35 (m, 2H), 3.15–2.9 (m, 2H), 2.05–1.8 (m, 2H), 1.8–1.35 (m, 3H), 1.02 (d, 3H). Anal. (C₁₆H₂₄ClN₃O₃·C₂H₂O₄·1.5H₂O).

2-(*cis*-3,5-Dimethylpiperidino)ethyl *N*-(4-amino-5-chloro-2-methoxyphenyl)carbamate (7g): oxalate salt (24%) from *i*-Pr₂O/MeOH; mp 136–138 °C; ¹H NMR (CD₃OD) δ 7.57 (s, 1H), 6.51 (s, 1H), 4.48 (m, 2H), 3.79 (s, 3H), 3.65–3.4 (m, 4H), 2.56 (m, 2H), 2.1–1.8 (m, 3H), 1.05–0.8 (m, 7H). Anal. (C₁₇H₂₆ClN₃O₃·C₂H₂O₄.H₂O).

2-Piperidinoethyl *N***-phenylcarbamate (8a):** hydrochloride salt (28%) from *i*-Pr₂O/MeOH; mp 216–218 °C; ¹H NMR (CD₃OD) δ 8.6 (s, 1H), 7.48 (m, 2H), 7.28 (m, 2H), 7.04 (m, 1H), 4.58 (m, 2H), 3.27 (m, 2H), 4–1.3 (m, 10H). Anal. (C₁₄H₂₁N₂O₂·HCl).

2-Piperidinoethyl N-(2-methoxyphenyl)carbamate (**8b**): oxalate salt (38%) from *i*- $Pr_2O/MeOH$; mp 180–182 °C; ¹H NMR (CD₃OD) δ 7.83 (m, 1H), 7.2–6.85 (m, 3H), 4.5 (m, 2H), 3.87 (s, 3H), 3.6–3 (m, 6H), 2–1.55 (m, 6H). Anal. (C₁₅H₂₂N₂O₃·C₂H₂O₄).

2-Piperidinoethyl *N***-(3-methoxyphenyl)carbamate (8d):** hydrochloride salt (43%) from *i*-Pr₂O/MeOH; mp 182 °C; ¹H NMR (CD₃OD) δ 7.19 (m, 1H), 7.13 (s, 1H), 6.97 (m, 1H), 6.61 (m, 1H), 4.5 (m, 2H), 3.76 (s, 3H), 3.46 (m, 2H), 3.9–2.8 (m, 4H), 2.1–1.55 (m, 6H). Anal. (C₁₅H₂₂N₂O₃·HCl).

2-Piperidinoethyl N-(4-methoxyphenyl)carbamate (8e): oxalate salt (56%) from *i*-Pr₂O/MeOH; mp 160 °C; ¹H NMR (CD₃OD) δ 7.33 (d, 2H), 6.84 (d, 2H), 4.45 (m, 2H), 3.75 (s, 3H), 3.41 (m, 2H), 3.7–2.9 (m, 4H), 2–1.45 (m, 6H). Anal. (C₁₅H₂₂N₂O₃·C₂O₄H₂).

2-Piperidinoethyl *N*-(5-chloro-2-methoxyphenyl)carbamate (8f): oxalate salt (47%) from *i*-Pr₂O/MeOH; mp 226– 228 °C; ¹H NMR (CD₃OD) δ 7.96 (d, 1H), 7.05 (dd, 1H), 6.96 (d, 1H), 4.51 (m, 2H), 3.87 (s, 3H), 3.44 (m, 2H), 3.5–3.15 (m, 4H), 1.96–1.6 (m, 6H). Anal. (C₁₅H₂₁ClN₂O₃·C₂O₄H₂).

2-Piperidinoethyl N-(2-chlorophenyl)carbamate (8g): hydrochloride salt (85%) from *i*-Pr₂O/EtOH; mp >220 °C dec; ¹H NMR (CD₃OD) δ 7.85 (m, 1H), 7.4 (m, 1H), 7.3 (m, 1H), 7.1 (m, 1H), 4.5 (m, 2H), 3.3 (m, 2H), 3.15 (m, 4H), 2.3 (m, 4H), 1.65 (m, 2H). Anal. (C₁₄H₁₉ClN₂O₂·HCl).

2-Piperidinoethyl N-(3-chlorophenyl)carbamate (**8h**): hydrochloride salt (84%) from *i*-Pr₂O/MeOH; mp 222 °C; ¹H NMR (CD₃OD) δ 7.6 (m, 1H), 7.31 (m, 1H), 7.25 (t, 1H), 7 (m, 1H), 4.5 (m, 2H), 3.45 (m, 2H), 3.9–2.7 (m, 4H), 2.1–1.4 (m, 6H). Anal. (C₁₄H₁₉ClN₂O₂·HCl).

2-Piperidinoethyl *N*-(**4-chlorophenyl**)**carbamate** (**8i**): hydrochloride salt (72%) from *i*-Pr₂O/EtOH; mp 214 °C; ¹H NMR (CD₃OD) δ 7.45 (d, 2H), 7.25 (d, 2H), 4.5 (m, 2H), 3.48 (m, 2H), 3.9–2.9 (m, 4H), 2.1–1.4 (m, 6H). Anal. (C₁₄H₁₉-ClN₂O₂·HCl).

2-Piperidinoethyl *N*-(2-aminophenyl)carbamate (8j): hydrochloride salt (77%) from *i*-Pr₂O/MeOH; mp 172 °C; ¹H NMR (CD₃OD) δ 7.15 (m, 1H), 7 (m, 1H), 6.82 (m, 1H), 6.69 (m, 1H), 4.46 (m, 2H), 3.6–3 (m, 6H), 2.1–1.5 (m, 6H). Anal. ($C_{14}H_{21}N_3O_2$ ·HCl).

2-Piperidinoethyl N-(3-aminophenyl)carbamate (**8k**): hydrochloride salt (88%) from *i*- $Pr_2O/MeOH$; mp 230 °C; ¹H NMR (DMSO) δ 10.8 (m, 1H), 10.16 (s, 1H), 7.59 (s, 1H), 7.42 (m, 1H), 7.35 (m, 1H), 7 (m, 1H), 4.5 (m, 2H), 3.46 (m, 2H), 3.35 (m, 2H), 2.97 (m, 2H), 2–1.5 (m, 5H), 1.3 (m, 1H). Anal. (C₁₄H₂₁N₃O₂·2HCl).

2-Piperidinoethyl *N*-(4-aminophenyl)carbamate (8): hydrochloride salt (34%) from *i*-Pr₂O/MeOH; mp >230 °C; ¹H NMR (DMSO) δ 10.55 (s, 1H), 10.06 (s, 1H), 7.55 (d, 2H), 7.31 (d, 2H), 4.48 (m, 2H), 3.46 (m, 2H), 3.34 (m, 2H), 2.95 (m, 2H), 2–1.6 (m, 5H), 1.77 (m, 1H). Anal. (C₁₄H₂₁N₃O₂·2HCl).

2-(*cis***-3,5-Dimethylpiperidino)ethyl N-(2-methoxy-phenyl)carbamate (8m):** oxalate salt (22%) from *i*-Pr₂O/ EtOH; mp 188–190 °C; ¹H NMR (CD₃OD) δ 7.83 (m, 1H), 7.1– 6.8 (m, 3H), 4.51 (m, 2H), 3.86 (s, 3H), 3.56 (m, 2H), 3.45 (m, 2H), 2.54 (t, 2H), 2.15–1.75 (m, 3H), 0.98 (d, 6H), 0.86 (m, 1H). Anal. (C₁₇H₂₆N₂O₃·C₂H₂O₄).

2-(*cis***-3,5-Dimethylpiperidino)ethyl** *N***-(***2***-ethoxyphen-yl)carbamate (8n):** oxalate salt (30%) from *i*-Pr₂O/MeOH; mp 164 °C; ¹H NMR (CD₃OD) δ 7.82 (m, 1H), 7.09–6.84 (m, 3H), 4.51 (m, 2H), 4.11 (q, 2H), 3.53 (m, 2H), 3.46 (m, 2H), 2.55 (t, 2H), 2.1–1.75 (m, 3H), 1.42 (t, 3H), 0.99 (d, 6H), 0.86 (m, 1H). Anal. (C₁₈H₂₈N₂O₃·C₂H₂O₄).

2-(*trans***-3,5-Dimethylpiperidino)ethyl N-(2-ethoxy-phenyl)carbamate (80):** oxalate salt (6%) from *i*-Pr₂O/MeOH; mp 174–176 °C; ¹H NMR (CD₃OD) δ 7.81 (m, 1H), 7.1–6.8 (m, 3H), 4.69–4.4 (m, 2H), 4.11 (q, 2H), 3.5–3.4 (m, 2H), 3.4–2.9 (m, 2H), 2.35–2.1 (m, 2H), 1.53 (m, 2H), 1.42 (t, 3H), 1.08 (d, 6H). Anal. (C₁₈H₂₈N₂O₃·C₂O₄H₂).

2-Piperidinoethyl *N*-(2-nitrophenyl)carbamate (8p): hydrochloride salt (83%) from *i*-Pr₂O/EtOH; mp 164 °C; ¹H NMR (CD₃OD) δ 8.2 (m, 1H), 8.12 (m, 1H), 7.68 (m, 1H), 7.27 (m, 1H), 4.58 (m, 2H), 3.51 (m, 2H), 3.8–2.8 (m, 4H), 2.2–1.4 (m, 6H). Anal. (C₁₄H₁₉N₃O₄·HCl).

2-Piperidinoethyl *N*-(3-nitrophenyl)carbamate (8q): hydrochloride salt (61%) from *i*- $Pr_2O/MeOH$; mp >210 °C; ¹H NMR (CD₃OD) δ 8.49 (m, 1H), 7.87 (m, 1H), 7.75 (m, 1H), 7.51 (m, 1H), 4.56 (m, 2H), 3.49 (m, 2H), 3.9–2.9 (m, 4H), 2.2–1.4 (m, 6H). Anal. (C₁₄H₁₉N₃O₄·HCl).

2-Piperidinoethyl *N*-(4-nitrophenyl)carbamate (8r): base (71%) from petroleum ether; mp 104 °C; ¹H NMR (CDCl₃) δ 8.15 (d, 2H), 7.5 (d, 2H), 7.45 (s, 1H), 4.43 (t, 2H), 2.65 (t, 2H), 2.45 (m, 4H), 1.7–1.3 (m, 6H). Anal. (C₁₄H₁₉N₃O₄).

3-Piperidinopropyl *N*-(2-methoxyphenyl)carbamate (8s): oxalate salt (34%) from *i*-Pr₂O/MeOH; mp 170 °C; ¹H NMR (CD₃OD) δ 7.82 (d, 1H), 6.81–7.1 (m, 3H), 4.22 (m, 2H), 3.86 (s, 3H), 3.8–3.35 (m, 2H), 3.27–3.15 (m, 2H), 3.15–2.6 (m, 2H), 2.3–2.23 (m, 2H), 1.6–1.4 (m, 6H). Anal. (C₁₆H₂₄N₂O₃·C₂H₂O₄).

1-(3-(1-Piperidinyl)propanecarboxamido)-4-amino-5chloro-2-methoxybenzene (9b). 4-Chlorobutyryl chloride (0.7 g, 5 mmol) in 10 mL of benzene was added to 1 g (5 mmol) of 5-chloro-4-nitro-2-anisidine in 20 mL of benzene at room temperature, and the solution was stirred for 30 min. Then 50 mL of a saturated aqueous solution of Na₂CO₃ was introduced, and the mixture was stirred for 15 min. The organic solution was separated, washed with a 1 N HCl aqueous solution and water, and dried over MgSO₄. The solvent was evaporated to give 1 g of chloroamide 15 (65%) which was added to a mixture of 4 mL of piperidine and toluene and refluxed for 4 h. The precipitate of piperidine hydrochloride was filtered, and the organic solution was evaporated. The crude product was dissolved in AcOEt, and the addition of 3 N HCl gave 9a hydrochloride (0.5 g) which was isolated by filtration as an orange powder. The evaporation of the filtrate provided 0.6 g of 1-(5-chloro-2-methoxy-4nitrophenyl)-2-pyrrolidone as the product of the cyclization reaction; 0.5 g of 9a hydrochloride was dissolved in a mixture of HOAc and concentrated HCl and stirred in the presence of SnCl₂ overnight. Then, it was poured onto ice, basified, and extracted by EtOAc. The organic solution was dried over MgSO₄ and evaporated. The crude material was transformed into the hydrochloride salt and recrystallized from a i-Pr₂O/ MeOH mixture to give 150 mg of 9b (33%): mp 190-200 °C;

¹H NMR (CD₃OD) δ 8.4 (s, 1H), 7.1 (s, 1H), 3.55 (m, 2H), 3.45 (s, 3H), 3.15 (m, 2H), 2.95 (m, 2H), 2.65 (m, 2H), 2.2-1.6 (m, 7H), 1.55 (m, 1H). Anal. $(C_{16}H_{24}ClN_3O_2 \cdot 2HCl \cdot 2H_2O)$

1-(3-(1-Piperidinyl)propanecarboxamido)-2-methoxybenzene (9c). A 3.4 mL aliquot (30 mmol) of 4-chlorobutyryl chloride in 50 mL of benzene was added to 4.9 g (40 mmol) of 2-anisidine in 100 mL of benzene at room temperature, and the solution was stirred for 30 min. Then 50 mL of a saturated Na₂CO₃ aqueous solution was added, and the mixture was stirred for 15 min. The organic solution was separated, washed with a 1 N HCl aqueous solution and water, and dried over MgSO₄; 6.8 g of crude product was obtained after the evaporation of the solvent. It was added to 10 mL of piperidine and stirred at room temperature in the presence of 0.1 g of NaI for 2 days. The piperidine was evaporated, and the residue was taken up in ether, a 10% Na₂CO₃ aqueous solution, and water, and dried over MgSO₄. After evaporation of the solvent, the crude compound was transformed into the hydrochloride salt with 3 N HCl/AcOEt solution and recrystallized from an AcOEt/EtOH mixture (7 g, 74.6%): mp >220 °C; 1 H NMR (CD₃OD) δ 7.9 (m, 1H), 7.15 (m, 1H), 7 (m, 1H), 6.9 (m, 1H), 3.85 (s, 3H), 3.1 (m, 4H), 3 (m, 2H), 2.6 (m, 2H), 2.05 (m, 2H), 1.8 (m, 4H), 1.62 (m, 2H). Anal. (C16H24N2O2·HCl).

Pharmacological Methods. 1. 5-HT₄ Receptor Binding Assays. Male Sprague-Dawley rats from Janvier Laboratories (France) were used. Animals were housed at 22 ± 1 °C, with 55% humidity, on a 12 h light–dark cycle with free access to food and water for 4 days before the experiments.

Membranes were prepared from rat striatum and olfactory tubercules which were pooled separately and stored at -80 °C. Tissues were thawed at 0 °C, homogenized in 15 volumes of ice-cold 50 mM Hepes (pH 7.4) using a Polytron homogenizer, and then centrifuged once at 40000g at 4 °C for 15 min. The resulting pellet was resuspended in 5 volumes of Hepes buffer to a final concentration of 4-5 mg of protein/mL. Membrane aliquots of 2.6 mL were kept frozen at -80 °C for subsequent use.

Binding assays were performed according to the method previously described¹² with modifications. The binding of [³H]-GR 113808 (85 Ci/mmol; Amersham) was measured using membranes (50 μ L aliquots, equivalent to 0.1–0.2 mg of protein) suspended in a final volume of 0.5 mL of 50 mM Hepes buffer (pH 7.4). Seven concentrations of each drug were used, and the assay was done in triplicate. Nonspecific binding was defined with 10 μ M ML 10302 in duplicate and represented less than 10% of the total binding. Total binding was defined in quadruplicate.

For each assay the bound radioactivity was separated by vacuum filtration through Whatman GF/B glass filters presoaked in 0.1% poly(ethylenimine) using a Brandel Cell harvester. The filters were then washed twice with 5 mL of 50 mM Tris-HCl (pH 8.4) at room temperature and dried. The filters were placed in polyethylene vials to which was added 4 mL of a scintillation cocktail (Beckman, Ready-Safe), and after equilibration, the radioactivity was determined using liquid scintillation spectrometry. The data were analyzed by a computer-assisted curve-fitting program in Lotus 1.2.3. to provide IC₅₀, K_i , and r^2 values, K_i values being calculated from the Cheng-Prussof equation.

2. Protein Estimation. The protein concentrations of the rat striatum and olfactory tubercule membrane preparations were determined by the method of Bradford.²⁰

3. 5-HT₄ Receptor Activity: Myenteric Plexus and Longitudinal Muscle Preparation from Guinea Pig Ileum. Tissues were obtained from male Crl (HA) SPF guinea pigs (400-600 g) housed as described previously. According to Craig and Clarke,^{4a} segments (2 cm long, 10 cm from the ileo-cecal junction) of myenteric plexus and longitudinal muscle from the ileum were carefully dissected and mounted in a 20 mL organ bath containing a warm (37 °C), aerated (95% O₂, 5% CO₂) Krebs solution. The tissue was stretched with a 0.5 g weight, and contractions elicited by transmural electrical stimulation (0.2 Hz, pulse duration of 1.5 ms) were recorded by means of an isotonic transducer connected to a polygraph (Gemini, Basile, Italy). The preparations were subjected to a supramaximal voltage stimulation. After stabilization and incubation with 10⁻⁷ M phenoxybenzamine, the voltage was reduced to obtain contractions 50% of those elicited by supramaximal stimulation. In control experiments, at least three fully reproducible cumulative concentration-effect curves to 5-HT (3 \times 10⁻¹⁰-10⁻⁷ M, contact time of about 60 s) were obtained. After two fully reproducible cumulative concentration-effect curves to 5-HT, a cumulative concentration-effect curve to the 5-HT₄ receptor agonist under test was carried out (contact time of 60-120 s). The agonist activity (EC₅₀, nM) was assessed as the concentration which gave a 50% increase in the response to the electrical stimulation with regard to the maximum, and the results are expressed as a percentage of the maximal effect of 5-HT (100%) found in the second reference curve. In antagonist studies, the compound under test was added 30 min before the last cumulative concentration–effect curve to 5-HT (3 \times $10^{-10}{-}10^{-5}$ M). Several concentrations ($n \ge 3$) of antagonist were tested, and the IC₅₀ value was calculated as the concentration producing a 50% reduction of the submaximal (80%) 5-HT contraction. The submaximal (80%) 5-HT concentration (3 \times 10 $^{-8}$ M) was calculated on the second reference cumulative concentrationresponse curve.

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